

Generation and validation of anti-linker monoclonal antibodies for the surface detection of scFv-based CARs

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INTRODUCTION

Chimeric Antigen Receptor (CAR)-T cell therapy is a highly innovative form of immunotherapy that has proven to be successful in the treatment of B cell malignancies and multiple myeloma. As this treatment modality continues to evolve toward the targeting of novel tumor antigens and engineering of cells with greater persistence, there is a need in multiple phases of the CAR-T development pipeline for highly specific detection reagents that can be leveraged to monitor the expression of CARs on the cell surface. Many commercially available CAR detection reagents, however, either lack specificity or are not versatile in their ability to detect CARs of differing antigen specificity. Here, we report on the generation and validation of rabbit monoclonal antibodies raised against two linker sequences that are commonly integrated into single-chain variable fragment (scFv)-based CARs. These antibodies serve as versatile detection reagents that can be used to interrogate the surface expression of CARs.

METHODS

The recombinant monoclonal antibodies, E7O2V and E3U7Q, were generated by rabbits immunized with peptide sequences most used for the linker region of scFv based CARs, Gly₄Ser and Whitlow, respectively (1,2). E7O2V and E3U7Q were directly conjugated to a panel of fluorophores and validated for specificity and versatility using live cell flow cytometric analysis of nontransduced versus CAR-transduced cells.

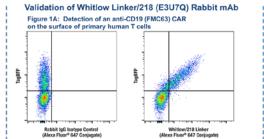
CONCLUSIONS

In a live cell flow cytometry assay, fluorophore conjugates of E7O2V and E3U7Q specifically detect surface expressed scFv-based CARs containing either a Gly₄Ser linker or a Whitlow linker, respectively. Furthermore, these recombinant monoclonal antibodies are versatile in that they can also detect their respective linker sequence independently of scFv specificity. The potential exists to leverage these antibodies for CAR-T cell enrichment and for incorporation into multiparametric flow cytometry panels used to phenotype CAR-T cells during the discovery, manufacturing, and clinical phases of the development pipeline.

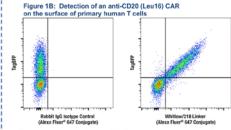
REFERENCES

- Huston, J.S. et al. (1988) Proc Natl Acad Sci U S A 85, 5879-83.
- 2. Whitlow, M. et al. (1993) Protein Eng 6, 989-95.

Staining Reagent	Limitations		
Anti-Idiotype	Limited to detection of CARs with a scFv of given specificity		
Protein L	 Differential binding to κ light chain variable regions Not amenable to incorporation into flow panels 		
Anti-Fab	 Non-specificity Not amenable to incorporation into flow panels 		
Anti-Fc	 Non-specificity Limited to detection of CARs with an Ig-derived hinge 		
CAR target antigen	Limited to detection of a CAR directed against a specific target antigen		
(E702V) Rai	bbit mAb (E3U7Q) Rabbit mAb		
CCCCSSCCCSSCCCCS	oblit mAb		
	COTICOSCICKPOSOECOSTIKO		
	COTICOSCICKPOSOECOSTIKO		
	COTICOSCICKPOSOECOSTIKO		

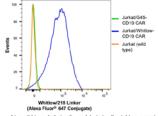


Flow cytometric analysis of live pan-C03+ T cells engineered to express a scFv-based Anti-CD19 (FMC63) CAR containing a Whitlow/218 linker, using Whitlow/218 Linker (E3UTQ) Rabit mA/ (Alexa Flucr® 447 Conjugate) (right) or concentration-matched Rabit (DA1E) mAb tgG XPP botype Control (Alexa Fluor® 647 Conjugate) #2586 (reft). Tag Blue fluorescent protein (TogBPP) is a transduction marker co-expressed with the CAR.

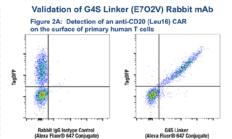


Flow cytometric analysis of live pan-CD3+T cells engineered to express a scFv-based Anti-CD20 (Leu16) CAR containing a Whitlow/218 linker, using Whitlow/218 Linker (E3U7Q) Rabbit mbb (Alexa Fluor6 647 Conjugate) (right) or concentration-matched Rabbit (DA1E) mbb (gG XP8 Isotype Control (Alexa Fluor6 647 Conjugate) #2985 (lett). Tag Blue fluorescent protein (riggJFP) is a transduction marker co-expressed with the CAR.

Figure 1C: Whitlow Linker/218 (E3U7Q) Rabbit mAb does not cross-react with a scFv-based CAR using a G4S linker

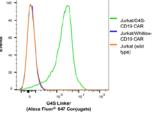


Live wild type Jurkat cells, and Jurkat cells stably expressing a scFv-based CD19 (FMC63) CAR using either a 3X G4S linker or a Whitlow/218 linker, were immunostained with Whitlow/ 218 Linker (E3U7O) Rabbi mkb (Alexa Fluor® 647 Conjigate) and analyzed by flow cytometry.



Flow cytometric analysis of live pan-CD3+ T cells engineered to express a scFv-based Anti-CD20 (Leu16) CAR containing a G4S linker, using G4S Linker (E7O2V) Rabbt mAb (Alexa Fluxe® G47 Conjugate) (right) or concentration-matched Rabbi (DA1E) mAb IgG XM® lostype Control (Alexa Fluxe® 647 Conjugate) #2955 (left). Tag Blue flucrescent protein (TagBFP) is a transduction marker co-expressed with the CAR.

Figure 2B: G4S Linker/218 (E7O2V) Rabbit mAb does not crossreact with a scFv-based CAR using a Whitlow/218 linker



Live wild type Jurkat cells, and Jurkat cells stably expressing a scFv-based CD19 (FMC63) CAR using either a 3X G4S linker or a Whitlow/218 linker, were immunostained with G4S Linker (E7O2V) Rabbit mAb (Alexa Fluor® 647 Conjugate) and analyzed by flow cytometry.

Table 2:	Selected example	of anti-CAR linker n	abbit mAb conjugates

Product Name	Catalog #
G4S Linker (E7O2V) Rabbit mAb (PE Conjugate)	#38907
G4S Linker (E7O2V) Rabbit mAb (Alexa Fluor® 647 Conjugate)	#69782
G4S Linker (E7O2V) Rabbit mAb (Alexa Fluor® 488 Conjugate)	#50515
G4S Linker (E7O2V) Rabbit mAb (Biotinylated)	#17621
Whitlow/218 Linker (E3U7Q) Rabbit mAb (PE Conjugate)	#62405
Whitlow/218 Linker (E3U7Q) Rabbit mAb (Alexa Fluor® 647 Conjugate)	#69310
Whitlow/218 Linker (E7O2V) Rabbit mAb (Alexa Fluor® 488 Conjugate)	#55809
Whitlow/218 Linker (E3U7Q) Rabbit mAb (Biotinylated)	#32523



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